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Rapid fabrication of microfluidic PDMS devices from reusable PDMS molds using laser ablation

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Abstract

The conventional fabrication methods for microfluidic devices require cleanroom processes that are costly and time-consuming. We present a novel, facile, and low-cost method for rapid fabrication of polydimethylsiloxane (PDMS) molds and devices. The method consists of three main fabrication steps: female mold (FM), male mold (MM), and chip fabrication. We use a CO₂ laser cutter to pattern a thin, spin-coated PDMS layer for FM fabrication. We then obtain reusable PDMS MM from the FM using PDMS/PDMS casting. Finally, a second casting step is used to replicate PDMS devices from the MM. Demolding of one PDMS layer from another is carried out without any potentially hazardous chemical surface treatment. We have successfully demonstrated that this novel method allows fabrication of microfluidic molds and devices with precise dimensions (thickness, width, length) using a single material, PDMS, which is very common across microfluidic laboratories. The whole process, from idea to device testing, can be completed in 1.5 h in a standard laboratory.

Keywords: microfabrication, microfluidics, rapid prototyping, laser ablation, microdroplet generation

 Online supplementary data available from stacks.iop.org/JMM/26/035008/mmedia

(Some figures may appear in colour only in the online journal)

1. Introduction

Microfluidics, the study and manipulation of fluids in micrometer scale structures, provides promising platforms for chemical and biological experimentation for lab-on-a-chip applications [1]. Polydimethylsiloxane (PDMS) is commonly used for microfluidic devices because it is optically transparent and biocompatible. Soft lithography is the most preferred method for the fabrication of PDMS devices [2]. This method employs a mold that can be used multiple times for device fabrication. In order to get the desired device patterns on the PDMS layer, one should fabricate the mold, cast

PDMS onto the mold, cure PDMS, and peel off the PDMS layer. Then, the device fabrication is completed by punching the inlet/outlet holes and bonding to another surface.

UV photolithography provides down to 1 μm feature resolution for molds used for soft lithography [3]. However, fabrication of the mold requires cleanroom processes that are expensive, time-consuming, and labour intensive. Also, commonly used SU-8/silicon molds are fragile and prone to delamination [4]. It is therefore of critical importance to develop alternative rapid prototyping methods. Direct writing of PMMA and PDMS via laser ablation has been previously demonstrated [5–9]. In these studies, bulges, splashes, and

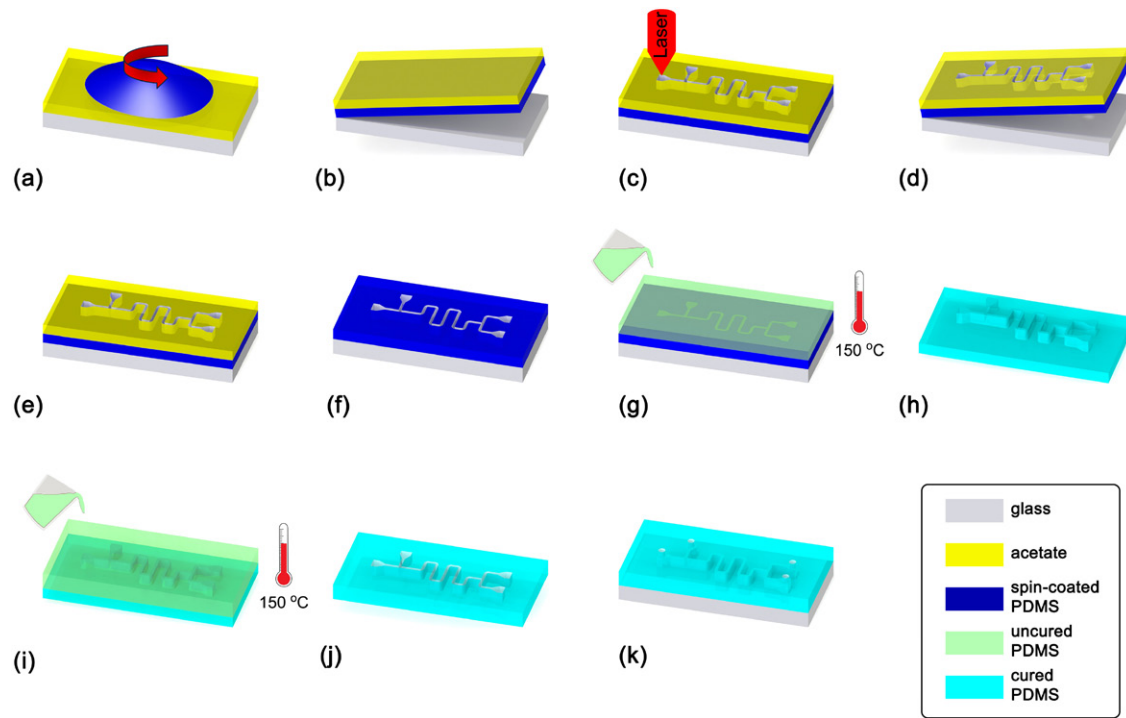


Figure 1. Rapid fabrication process flow for PDMS microfluidic devices. (a) PDMS is spin-coated on a 150 μm thick acetate sheet. (b) The PDMS/acetate layers are flipped over on the glass slide. (c) The PDMS/acetate layers are patterned by laser ablation. (d) The glass slide is discarded, and the PDMS/acetate layers are washed with DI. (e) The PDMS/acetate layers are plasma-bonded on a glass slide. (f) The acetate layer is peeled off to form the female mold. (g) PDMS is cast on the female mold and cured for 2 min at 150 $^{\circ}\text{C}$. (h) The top PDMS layer is peeled off to form the male mold. (i) PDMS is cast on the male mold and cured for 2 min at 150 $^{\circ}\text{C}$. (j) The top PDMS layer is peeled off to form the chip. (k) The inlet/outlet holes are punched, and the chip is plasma-bonded on a glass slide to form the microfluidic device.

resolidification of ablated material cause distorted sidewalls and bonding issues. Also, the channel height is dependent on laser parameters that are prone to change over long runs. Additionally, in direct writing, the whole process must be repeated each time from scratch to get a new device. To address the shortcomings of direct writing and fragility of silicon molds, elastomer (PDMS) and plastic molds have been fabricated using replica molding [4, 10]. However, these molds require cleanroom-fabricated silicon molds to start with. Also, for PDMS molds, adhesion occurs during PDMS/PDMS casting [11, 12]. To prevent adhesion, several chemical surface treatments have been demonstrated with potentially hazardous chemicals [13–16].

In this study, we introduce a novel rapid fabrication method to fabricate PDMS microfluidic molds and devices. We can obtain channels with precise dimensions and vertical side walls, which enables this method to be an alternative for cleanroom-dependent fabrication methods. This method provides significant time and cost savings especially when several design iterations are required. The method has three main steps. In the first step, we fabricate the female mold (FM) by patterning a thin spin-coated PDMS layer using a CO_2 laser cutter. We can precisely control the thickness of the PDMS layer through spin coating settings, which in turn determines the channel height. In the second step, we obtain the reusable PDMS male mold (MM) from the FM by replica molding. This MM replaces the silicon mold in soft lithography. Finally, we obtain PDMS devices from the reusable MM by PDMS/PDMS casting.

We have demonstrated that demolding of PDMS devices from a PDMS mold can simply be performed by applying a drop of silicone oil between the PDMS layers and by carefully setting the curing time and temperature. To demonstrate the longevity of the reusable PDMS MM, we compared the first and the tenth replicas obtained from the same mold. Also, in order to demonstrate the versatility of the method, we fabricated a T-junction microfluidic device and showed formation of monodisperse water-in-oil microdroplets.

2. Materials and methods

The rapid fabrication process of PDMS microfluidic devices is schematically summarized in figure 1. One of the key benefits of this method is that both the FM and the MM are fabricated from PDMS (Sylgard 184, Dow Corning), which is a standard material used in microfluidic laboratories. The fabrication process consists of three main steps: (i) FM fabrication using laser ablation, (ii) MM fabrication using PDMS/PDMS casting, and (iii) device fabrication using PDMS/PDMS casting.

2.1. Fabrication of the female mold

For the fabrication of the FM, firstly, a 150 μm thick acetate sheet is taped on a glass slide. Then, PDMS (prepared at 10 : 1 ratio, debubbled) is spin-coated on the acetate sheet (figure 1(a)). The thickness of the PDMS layer can be precisely controlled

through spin coating settings, which define the channel height [17]. For the spin coating parameters for different channel heights, see figure S1 (stacks.iop.org/JMM/26/035008/mmedia) on supplementary data file.

Following spin coating, the PDMS layer is cured for 20 min on a hotplate at 100 °C. After curing the thin PDMS layer, the PDMS/acetate structure is removed from the glass slide and flipped over on a clean glass slide such that the PDMS layer is sandwiched between the acetate sheet and the glass slide (figure 1(b)). Then, the spin-coated PDMS layer and the acetate sheet are laser-ablated to form the desired microfluidic channel pattern that is designed using a CAD software (figure 1(c)). It is demonstrated in the literature that upon laser ablation, there remain some bulges and debris around the ablated areas due to back condensation of the ablated material [5, 9, 18, 19]. These leftover pieces deteriorate the molding process in the subsequent steps. To avoid such issues, it is important to protect the surface of the spin-coated PDMS layer. Hence, we form the glass/PDMS/acetate structure before laser ablation. The acetate layer protects the PDMS surface whereas the glass slide provides the structural support. A comparison of the laser ablation results with and without the acetate layer is provided in the supplementary data file.

We use a 30W CO₂ laser (Epilog, Zing) to transfer the desired patterns onto the FM. Most laser cutters have two operation modes: raster and vector. We have found that the vector mode results in less debris formation and smoother channel sidewalls, thus provides better ablation performance for PDMS. The power, frequency, and speed of the laser cutter are set as 6W, 1000 Hz, and 2.5 cm s⁻¹, respectively. During the optimization of the laser parameters, we have scanned a range of values for these parameters and evaluated the outcome in terms of the channel sidewall profile, the precision of channel width, and the amount of debris formed. We have seen that laser power less than 4.5W does not ablate the PDMS/acetate layers. If the laser power is above 6.5W, it burns the PDMS layer and leaves ash on the surface. The device patterns are designed in 2D using a CAD software (Autodesk, AutoCAD 2015). The channel width is defined by both the 2D drawing and the laser spot size. The focused spot size of the laser beam for our system is 120 μm. If the desired channel width is smaller than 120 μm, the spot size can be reduced by additional commercial or custom-made optical components [20, 21]. In case the desired channel width is larger than 120 μm, the laser can be traced along the channel walls. During channel fabrication by laser ablation, the laser beam ablates the channel intersection points more than once. In conventional laser writing techniques, this causes over ablation and results in a deeper channel at the channel junction as seen in several studies in the literature [9, 22]. On the other hand, we are making a through cut on the thin, spin-coated PDMS layer by laser ablation. Therefore, even though the same location is traced by the laser beam multiple times, the ablated thickness does not vary. This method allows very precise control of channel dimensions as opposed to the previously demonstrated laser-ablated devices.

The formation of glass/PDMS/acetate structure is also of importance to provide the draft angle required during the

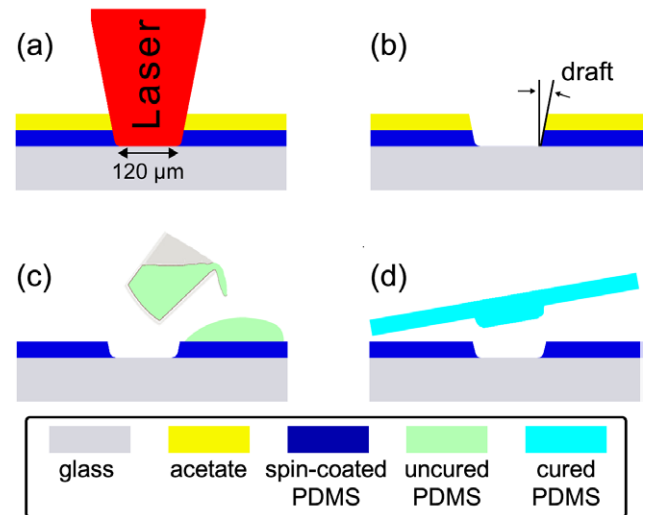


Figure 2. Schematic representation of the formation of the draft angle with laser ablation. (a) The PDMS/acetate layers are patterned by laser ablation figure 1(c). (b) The patterned layers are plasma-bonded on a glass slide figure 1(e), which forms a positive draft angle. (c) PDMS is cast on the FM figure 1(g). (d) The cured PDMS layer is peeled off figure 1(h). The draft allows easier peeling-off.

subsequent casting steps. The side view of the laser ablation step (figure 1(c)) is shown in figure 2(a). The laser beam is focused on the glass slide so that the enlarging beam waist provides the draft angle. This tapered laser cut is the signature for laser ablation [7, 9]. When the laser-ablated PDMS layer is bonded to a separate glass slide (figures 1(e) and 2(b)), the orientation of the PDMS layer should be kept the same to have a positive draft angle. This ensures high replication fidelity for micrometer size channel designs.

Following the laser ablation step, the PDMS/acetate structure is peeled off from the glass slide and washed with deionized water to remove the debris originated from the laser-cutting step figure 1(d). Then, the laser-ablated PDMS layer is bonded to another glass slide with plasma activation figure 1(e). After plasma bonding, the acetate sheet is peeled off from the PDMS layer to form the FM figure 1(f). This FM is to be used for PDMS/PDMS casting to form the MM.

2.2. Fabrication of the reusable male mold

The second step in the fabrication process is the casting of the reusable PDMS MM. PDMS/PDMS double casting has been previously demonstrated in the literature [14, 23, 24]. However, for successful demolding, these studies require surface treatment using various chemicals [12, 14, 15, 25, 26]. Instead, we demonstrate that PDMS layers can simply be demolded from the bottom PDMS layer by applying silicone oil between the layers and suitably setting the curing time and temperature. Oil is absorbed by the bottom PDMS layer and prevents the adhesion of the PDMS layers, allowing facile release of one from the other without any deformation.

We start off by putting the FM in an aluminum pan with the laser-ablated PDMS layer facing up. Then, 100 μl silicone oil was dispensed over the PDMS using a pipette. 100 μl was enough for all the designs that we have used during this study.

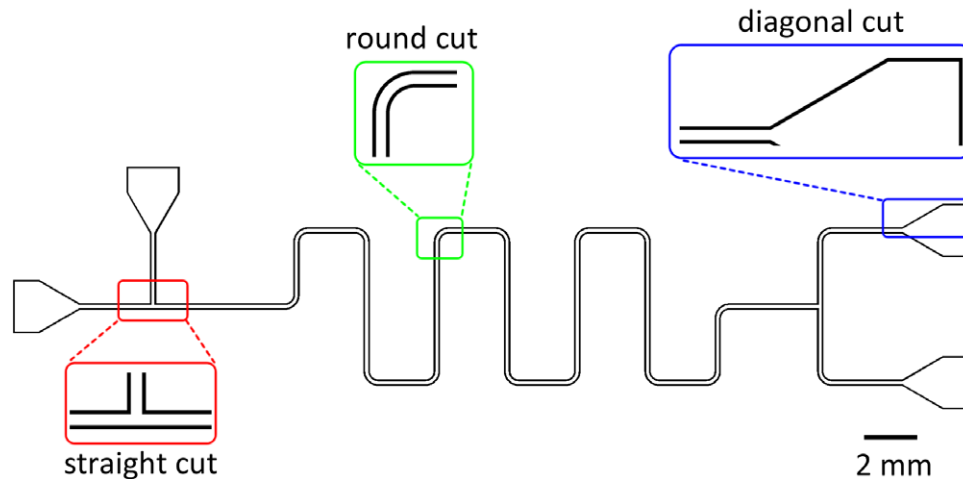


Figure 3. Schematic representation of the model microfluidic device designed on a CAD software (Autodesk, AutoCAD 2015). Three selected regions of interest are straight cut, round cut, and diagonal cut channel segments.

If the footprint of the chip is larger, more silicone oil can be dispensed so that all the recessed part in the FM is wet by oil. The excess amount of silicone oil spreads throughout the top layer. It is advised to cover the whole surface of the mold by oil. We wait for 1 min for the PDMS layer to absorb the silicone oil. Upon absorption, the excess oil is cleaned off the channels and the surface using an air gun. Then, the pan is placed on the hotplate for 5 min at 150 °C. While the pan is on the hotplate, PDMS (prepared at 10 : 1 ratio, debubbled) is poured on the FM figure 1(g). After 2 min, the pan is taken off the hotplate and cooled down to room temperature. It is critical to ensure that PDMS is hardened before taking the pan off the hotplate. Our experiments have demonstrated that it takes 2 min to harden 1-cm thick PDMS at 150 °C. Then, the PDMS MM is gently peeled off from the FM, and the negative copy of the desired device pattern is obtained as MM figure 1(h).

2.3. Fabrication of the chip

The MM obtained in the previous step can be used multiple times for chip fabrication. In order to fabricate the chip, we put the MM in an aluminum pan. Then, the pan is placed on the hotplate for 5 min at 150 °C. While the pan is on the hotplate, PDMS (prepared at 10 : 1 ratio, debubbled) is poured on the MM figure 1(i). After 2 min, the pan is taken off the hotplate and cooled down to room temperature. Then, the PDMS chip is gently peeled off from the MM figure 1(j). Finally, the inlet/outlet holes are punched, and the PDMS chip is bonded to a glass slide with plasma activation figure 1(k).

3. Results and discussion

3.1. Fabrication of a model device

To demonstrate the functionality of this method, we have designed a model T-junction droplet generator and sorter microfluidic device as shown in figure 3. The channel width was drawn as 300 μm . First, FM has been fabricated using the parameters given in the experimental section. It is important to note that when the vector laser-cutting mode is used, the laser

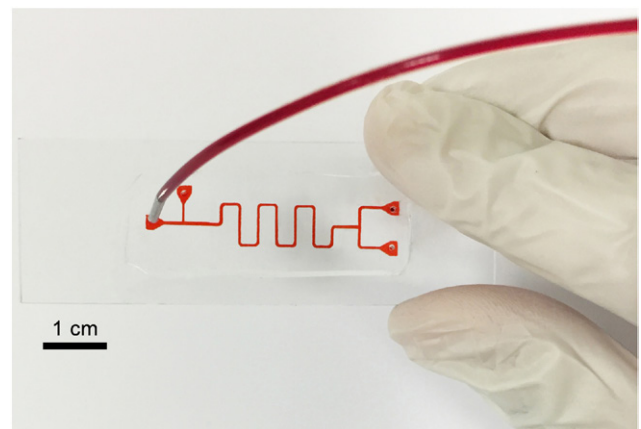


Figure 4. Image of the microfluidic device fabricated with the presented rapid fabrication method. The device is a T-junction droplet generator and sorter.

beam traces the channel walls throughout the design. Since the focused laser beam has a diameter of 120 μm , the final channel width obtained on the FM is higher than the designed width. In our case, we have measured the channel width as 450 μm . On the other hand, the channel depth is solely controlled by the spin-coating parameters of the PDMS layer that is patterned by the laser. When 10 : 1 mixed PDMS is spin-coated at a rate of 2000 rpm for 30 s, the PDMS thickness is 35 μm . After we fabricate the FM, we have obtained a reusable MM from the FM, and a chip from the MM, respectively.

We have defined three regions of interest on the model device to investigate the channel wall profile and the replication fidelity. The selected regions are composed of a straight, round, and diagonal cut channel segments. Figure 4 shows the final fabricated device.

Figure 5 shows the SEM images of these three regions of interest in the FM, MM, and chip. It should be noted that the laser cutter moves in stepwise motion in 2D plane. For this reason, round and diagonal patterns cannot be obtained as smooth as straight patterns. This results in around 5 μm roughness at the sidewalls. It should also be noted that the roughness of the channel sidewalls depends on the resolution

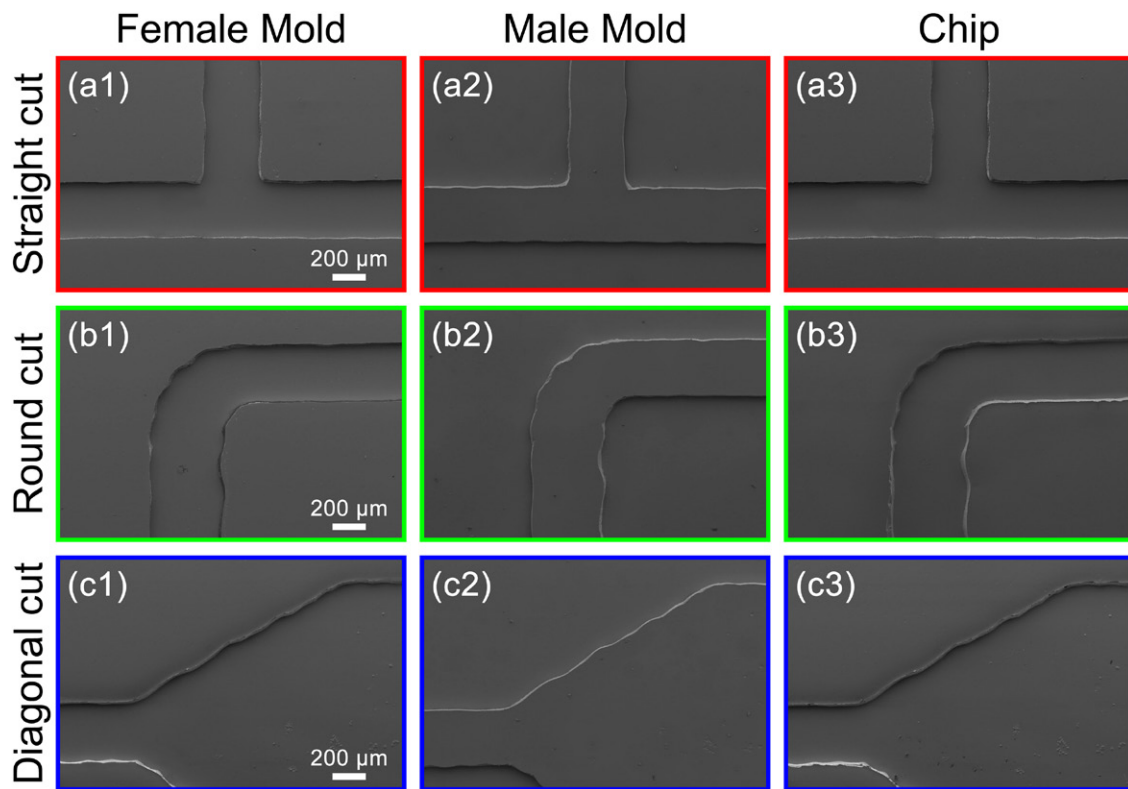


Figure 5. SEM images of the selected regions of the fabricated FM, MM, and chip. The first, second, and third rows give the images of the straight, round and diagonal cut regions, respectively. The left, middle, and right columns are the images of the FM, MM, and chip, respectively. The SEM images of the MM (a2, b2, c2) are mirrored for ease of comparison.

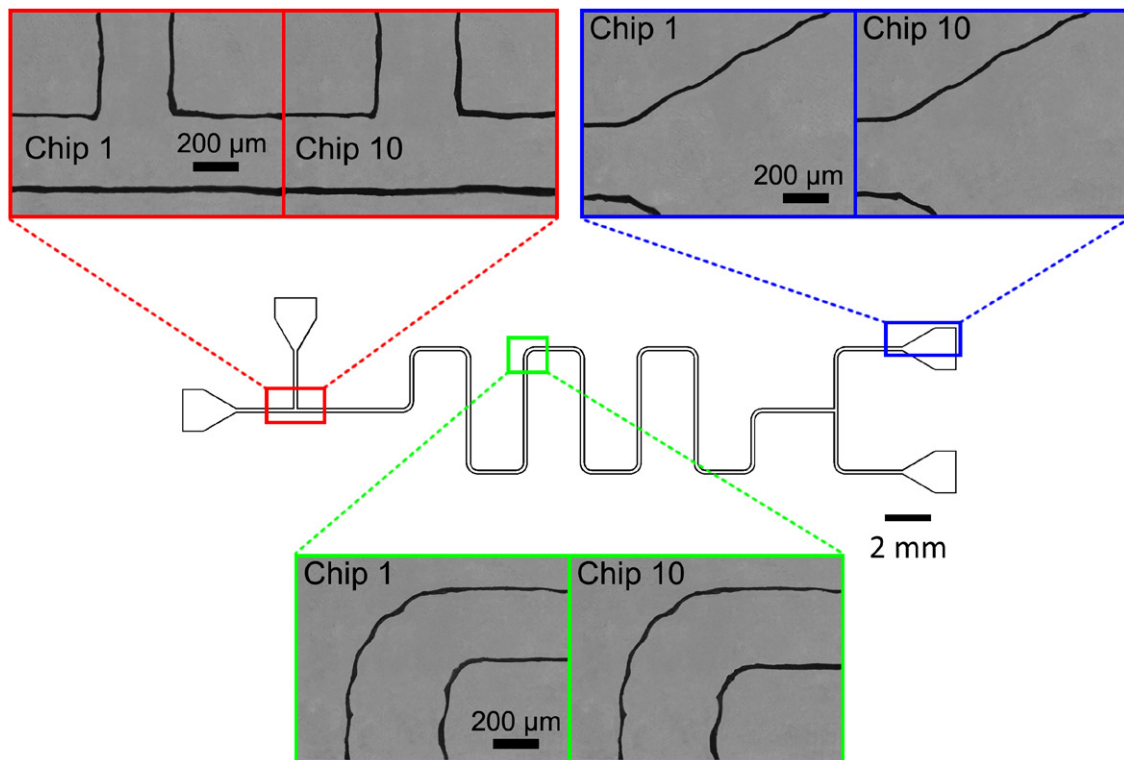


Figure 6. Optical microscope images of the selected regions of the first and tenth chips obtained from the same PDMS reusable MM. The comparison demonstrates that the fidelity performance of the method is very good.

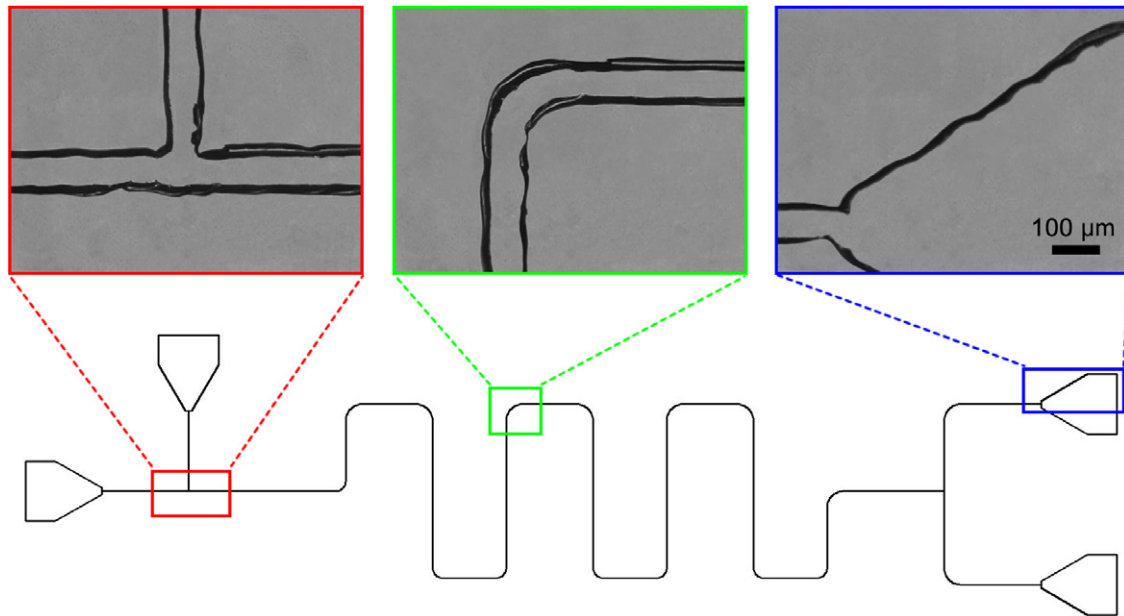


Figure 7. Optical microscope images of the selected regions of the chip with the smallest possible channel width, $120\ \mu\text{m}$. The chip is obtained from a FM formed by a single trace of laser beam.

of the laser cutter. For the laser parameters (speed and frequency) that we set, the roughness at the straight channel is around $1\ \mu\text{m}$. Two critical observations are made here. First, SEM images of the FM (figures 5(a1), (b1) and (c1)) prove that we can successfully obtain the desired vertical channel sidewalls throughout the device. We are able to ablate a thin PDMS layer without facing any of the laser ablation issues mentioned in the literature [9, 18]. Second, it is clear that both the MM and the chip replicate very well from the FM and the MM, respectively. These results show that PDMS/PDMS casting and demolding using silicone oil work successfully.

3.2. PDMS/PDMS replication fidelity

The main benefit of the SU-8 master molds fabricated using UV photolithography is their reusability. Therefore, it is important to evaluate the reusability of the molds produced by alternative fabrication methods. To demonstrate the replication fidelity of our method, we have obtained ten chips using the same PDMS MM. Figure 6 shows the optical microscope images of the previously discussed straight, round and diagonal sections of the first and the tenth chips. When we compare the channel dimensions for these two chips, we can see that the channel width varies less than 1%. This demonstrates that the very same MM can be used many times to get PDMS chips without any deformation.

Using the same fabrication method, we have also fabricated the same channel geometry with the smallest possible channel width, which is $120\ \mu\text{m}$ for our system. In this case, the CAD drawing of the design is modified, and the FM was formed by a single trace of laser beam. The 2D CAD drawing and the optical images of the chip are shown in figure 7. As seen, the channel width is approximately $120\ \mu\text{m}$; straight, round and diagonal cuts can be formed successfully. More importantly, the channel depth is the same throughout the design, since it

is dictated solely by the thickness of the PDMS spin-coated at the beginning of the fabrication. For the other laser ablation based methods, the junction regions, where two channels meet, have a higher depth since it is traced twice by the laser beam [9, 22]. On the other hand, our fabrication method completely alleviates this problem.

3.3. Microdroplet generation using the model device

To show the versatility of our fabrication method, we have generated monodisperse water droplets in silicone oil ($\mu = 100\ \text{mPa s}$, Ultrakim) using a T-junction microfluidic device with the design shown in figure 3. The inlet pressures for the oil and water were controlled using a pressure pump system (Elveflow OB1, 2 bar). Figure 8(a) shows the successive images of the formation of a water droplet, when the inlet pressures were set to 15 mbar for oil and 12 mbar for water inlets. At these pressures, the flow rates for oil and water were $0.0165\ \mu\text{l min}^{-1}$ and $0.006\ \mu\text{l min}^{-1}$, respectively. The droplet formation takes place in the squeezing regime [27, 28]. To analyze the size distribution of the generated water droplets, the length (front to end) of 100 subsequently generated droplets were measured using image processing (MathWorks, Matlab R2014). Figure 8(b) shows the histogram of the droplet sizes that yield a Gaussian distribution. The coefficient of variation for droplet size, which is defined as the ratio of standard deviation to mean, is calculated as 0.0346. This value is in accordance with the droplet monodispersity values for T-junction droplet generators in the literature [29, 30].

A wide range of microfluidic devices can be fabricated using our rapid fabrication method. We would like to emphasize that for any microfluidic device fabrication, the whole process, from idea to device testing, can be completed in 1.5 h as summarized in figure 9. In addition, the process does not require any complicated equipment and can be easily

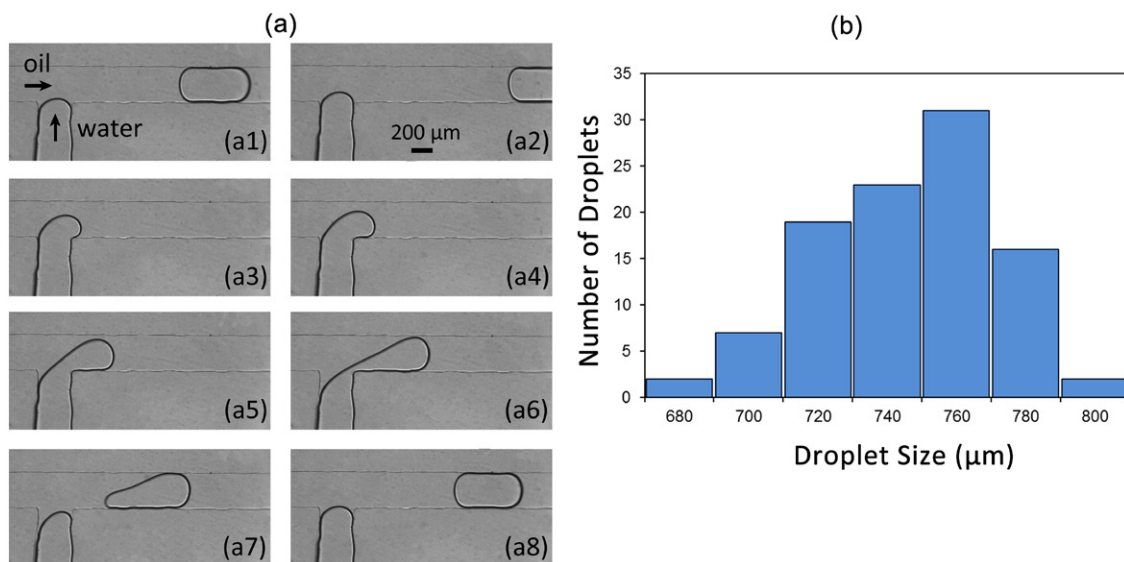


Figure 8. Monodisperse microdroplet generation using the fabricated model device. (a) Optical microscopy images (Omano OMFL600) of the generation of a water droplet in silicone oil in a time sequence. (b) Histogram showing the size distribution of 100 subsequently generated water droplets.

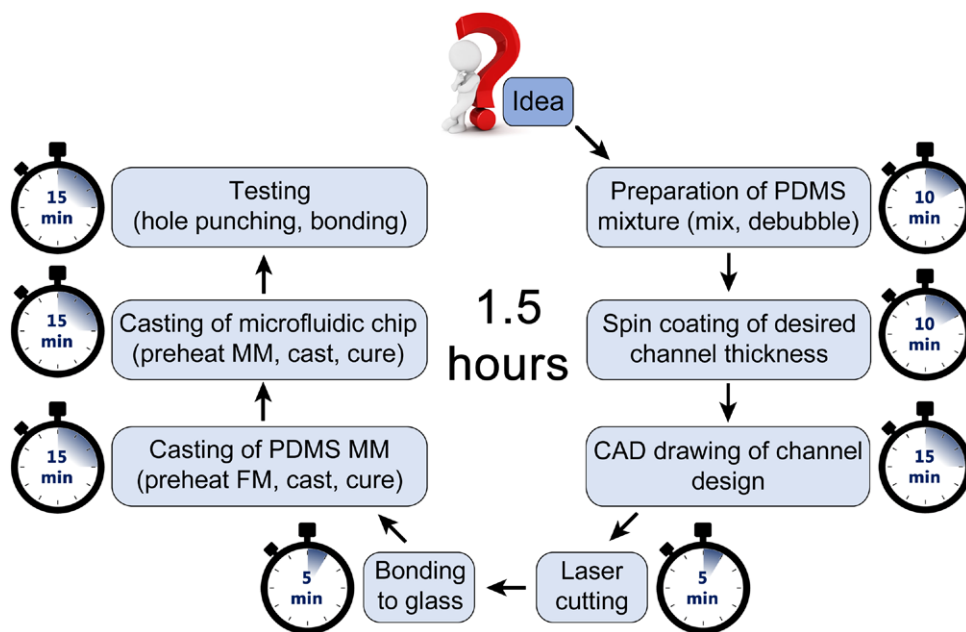


Figure 9. Workflow of the rapid fabrication method. From idea to device testing, the whole process can be completed in 1.5 h in a standard laboratory.

performed by a non-expert. The only limitation of this method is that the smallest channel width that can be obtained is determined by the spot size of the laser beam. Therefore, if the required channel width is smaller than the laser spot size of commercial laser cutter systems, additional optical components may be needed.

4. Conclusion

In this study, we have presented a facile and low-cost method for rapid fabrication of PDMS devices. We have used a laser cutter to pattern the channel design on a spin-coated PDMS

layer. Then, using two successive PDMS/PDMS casting steps, we have obtained the microfluidic devices. PDMS/PDMS casting can be performed without any chemical surface treatment. Once the MM is fabricated, it can be reused for chip fabrication similar to the silicon/SU-8 molds used in conventional soft lithography. To demonstrate the functionality of the method, we fabricated a model T-junction microdroplet generator device. The entire fabrication process was completed in a standard laboratory, and the time required from idea to device testing was 1.5 h. We believe this method will be especially useful when researchers need multiple channel design iterations for their studies.

Acknowledgments

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