

Potential protective effect of N-acetyl cysteine in acoustic trauma: An experimental study using scanning electron microscopy

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Abstract

Background. Oxidative stress has been associated with pathological processes involved in acoustic trauma.

Objectives. In this prospective experimental study, we investigated the potential preventive effect of N-acetyl cysteine (NAC) in rats exposed to acoustic trauma (AT). Light microscopic and scanning electron microscopic (SEM) evaluations were performed.

Material and methods. Healthy Wistar albino rats (n = 18) were divided into 3 groups: group 1 (control group, n = 6), group 2 (acoustic trauma group, n = 6), and group 3 (AT+NAC group, n = 6). The rats in group 2 were exposed to AT. The rats in group 3 received NAC at a dose of 100 mg/kg/day by gavage for 7 days, and then 10 min after the 7th-day dose, they were exposed to AT.

Results. From light and scanning electron microscopy evaluations in the control group, the cochlear structure and epithelium were normal. In group 2 (AT group), extensive hair cell loss was observed in the cochlea by light microscopy evaluation. In the SEM evaluation, various epithelial damage and loss of stereocilia were also observed. In group 3 (AT+NAC group), decreased damage with preserved cochlear structures was seen by light microscopy. In the SEM evaluation, although stereocilia loss was also seen, nearly normal cell structures and vertical and symmetrical alignment of stereocilia structures were observed compared to the AT group.

Conclusions. NAC reduced cochlear damage due to acoustic trauma. Because NAC has antioxidant capacity, AT may have caused an increase in free radicals and death of outer hair cells. NAC is an antioxidant agent and it prevented cochlear damage due to AT in rats.

Key words: N-acetyl cysteine (NAC), acoustic trauma (AT), light microscopy, scanning electron microscopy (SEM)

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Oxidative stress has been associated with pathological processes involved in auditory trauma including mitochondrial injury, activation of cell death pathways, activation of mediators of inflammation, glutamate excitotoxicity, and increased levels of lipid peroxidase.^{1–3} These findings suggest that antioxidants have the potential to block molecular cascades that are triggered by auditory trauma, which induces oxidative stress and results in permanent threshold shifts (PTS) and hearing loss.⁴

In blast-exposed ears, cochlear pathology includes scar formation replacing dead hair cells, fused and damaged stereocilia, and in some extreme cases, separation of the organ of Corti from the basilar membrane. Most of the hair cell loss was in the outer hair cell (OHC) region and generally in the middle or basal turn. Although hair cell loss was found in the apical turn, the damage was less pronounced. N-acetyl cysteine (NAC), which was used after blast exposure, significantly reduced the permanent structural and functional damage resulting from exposure to blast overpressure. Drug treatment suppressed both functional and physical damage to the OHCs.⁴

In the present study, we investigated the potential preventative effect of NAC in rats exposed to acoustic trauma (AT). In this experimental study, light microscopy and scanning electron microscopy (SEM) evaluations were performed.

Material and methods

The study occurred at Eskişehir Osmangazi University, Faculty of Medicine. The animals were maintained at the Experimental Animal Breeding and Experimental Studies Centre of Eskişehir Osmangazi University. Adaptation

and care of the animals and experimental studies were performed at the same center in compliance with the principles of the Declaration of Helsinki.⁵ Ethics committee approval was obtained from Eskişehir Osmangazi University.

Animal subjects

The study was performed in 18 healthy Wistar albino rats, weighing 190–220 g. Each group of rats was housed separately in a climate-controlled room at 20°C. The animals were randomly divided into 3 groups: group 1 (control; n = 6): rats in this group received no drug and were not subjected to AT. Group 2 (n = 6): rats in this group received no drugs and were exposed to AT. Group 3 (n = 6): rats in this group received NAC dissolved in distilled water at a dose of 100 mg/kg/day via gavage for 7 days. Then, 10 min after the 7th-day dose, they were exposed to AT (NAC+AT).

Anesthesia procedure

The rats were anesthetized during experiments via intramuscular injection of 40 mg/kg ketamine hydrochloride (Ketalar, Parke-Davis, USA), and 5 mg/kg xylazine hydrochloride (Rompun, Bayer, Germany). Eye-blink reflexes and respiratory rhythms were followed during the experiments, and deep anesthesia was achieved by repeated doses.

Acoustic trauma (AT)

To create AT in rats with 1–12 kHz band white noise, a MATLAB program that produced a single unit of variance sound was used. Then, the sound was filtered with

Fig. 1. Light microscopy views of cochlea in the control group, acoustic trauma group, and acoustic trauma+NAC groups. a. Control group: normal-appearing cochlea structure; b. Acoustic trauma group: extensive hair cell loss (double arrow), c. Acoustic trauma+NAC group: decreased damage with preserved cochlear structures was seen (sv – scala vestibuli, sm – scala media, st – scala tympani, vestibular membrane [▶], the organ of Corti [→]) (scale bar: 500 µm (a), scale bar: 200 µm (b), scale bar: 500 µm (c), hematoxylin and eosin (H&E) staining)

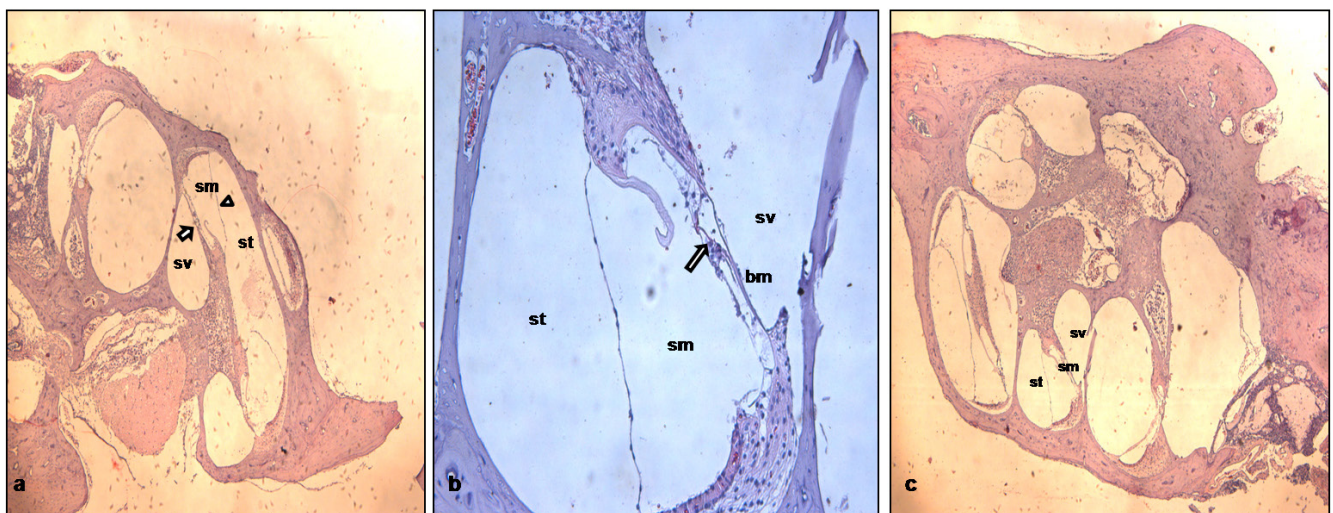
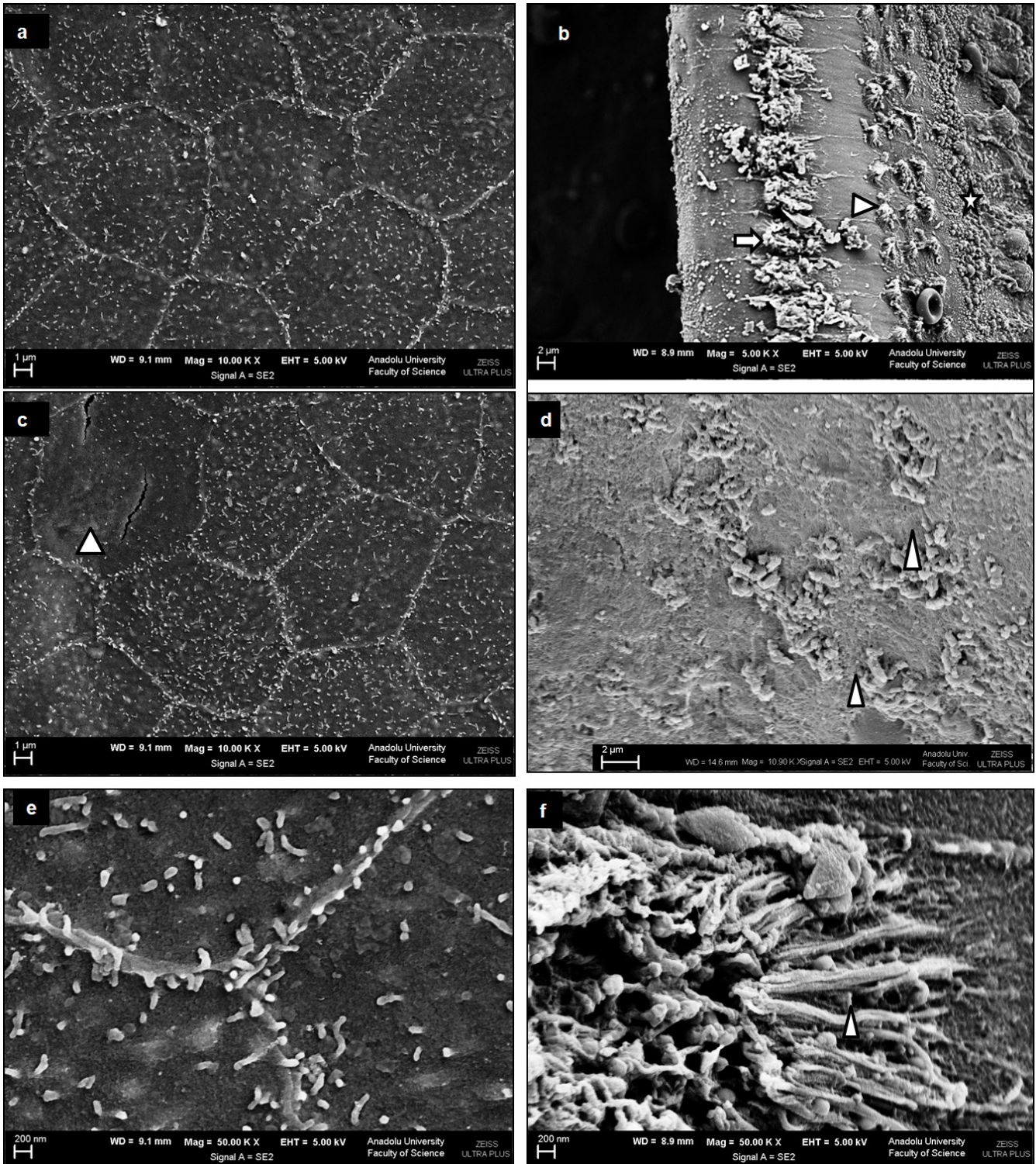


Fig. 2. Scanning electron microscopic (SEM) views of cochlea in the control group, acoustic trauma group, and acoustic trauma+NAC groups. a, b. Control group: in the control group, the SEM view of the epithelium showed a normal structure (a), with outer hair cell (▶), inner hair cells (→) and Hensen’s cells (*) (b); c, d. Acoustic trauma group: in cross-sectional images, various epithelial damage and loss of stereocilia were observed (▶) (c, d); e, f. Acoustic trauma+NAC group: although stereocilia loss was seen, nearly normal cell structure and vertical and symmetrical alignment of stereocilia structures were also observed, vs the acoustic trauma group (▶) (e, f) (SEM)



a 1–12 kHz FIR type digital filter, and an additional filter at 200 Hz. Then, the filtered noise was recorded in a computer-based Wav file. Set with a decibel meter, the AT was 110 dB of noise for 6 h (continuous noise).

Electron microscopy evaluation

After the aforementioned procedures were performed, the rats were sacrificed using 80 mg/kg pentothal. The time of sacrifice was 30 min after AT. Immediately after death, the temporal bones were removed, and the otic bullas were excised.⁶ Under a dissecting microscope, the bony capsule of the cochlea was carefully removed, and then the lateral wall was cut away to reveal the organ of Corti. We analyzed hair cell damage in 2 different areas of the middle and basal turns.

Scanning electron microscopy analysis

The cochlea and organ of Corti were dissected out and fixed for SEM preparation; all samples were fixed and post-fixed in the same way. Then, the cochlea was dehydrated through a graded ethanol series, prior to desiccation, using a critical-point drying method described by Lovell et al.⁷ Fully desiccated samples were mounted on a specimen stub using a carbon tab. After critical-point drying using CO₂ and sputter-coating with gold according to standard procedures, specimens were investigated with a Zeiss Ultra 50 SEM operating at 5 kV accelerating voltage.⁷

Results

Light microscopy and SEM evaluations showed that there was no cochlear damage in group 1 (control group). In group 2 (AT-exposed group), cochlear damage was evident. Inner hair cell stereocilia were missing in some regions, and OHC stereocilia were severely damaged by the noise trauma, with a large number prostrate or missing. The damage was more evident in the OHCs in the basal turns of the cochlea. In group 3 (NAC+AT group), although there was cochlear damage, it was less than in group 2 (AT-exposed group without NAC). In group 3, there were fewer missing cells in the inner and OHC group. There was slight damage to the OHC stereocilia.

Light microscopic evaluation results of groups 1–3 (Fig. 1)

Control group: Normal-appearing cochlear structures were observed (Fig. 1a). Acoustic trauma group: Extensive hair cell loss was detected in the cochlea (Fig. 1b). Acoustic trauma+NAC group: Decreased damage with preserved cochlear structures was seen (Fig. 1c).

Scanning electron microscopy evaluation results of Groups 1–3 (Fig. 2)

Control group: In the control group, SEM views of epithelium showed normal structures (Fig. 2a), outer hair cells, inner hair cells, and Hensen's cells (Fig. 2b). Acoustic trauma group: In cross-sectional images of this group, various epithelial damage and loss of stereocilia were observed (Fig. 2c, d). Acoustic trauma+NAC group: Although stereocilia loss was also present in this group, nearly normal cell structures and vertical and symmetrical alignment of stereocilia structures were also observed versus the acoustic trauma group (Fig. 2e, f).

Discussion

Acoustic overstimulation of the cochlea generates free radicals and reactive oxygen species (ROS), which may induce OHC death.^{8–10} Elevated levels of ROS activate the upregulation of cochlear antioxidant enzyme activity and modulate the key antioxidant compound, GSH.^{11–13} Various agents with antioxidant properties have been shown to attenuate threshold shifts and/or hair cell loss after the exposure to damaging noise.¹⁴

Oxidative stress plays a crucial role in the pathogenesis of NIHL and NIT. During exposure to excessive noise, the OHCs suffer metabolic depletion, leading to the accumulation of ROS and reactive nitrogen species (RNS), which may ultimately lead to necrosis and/or apoptosis.¹⁵ While necrosis is a passive form of cell death, usually occurring after gross physical or chemical insult, associated with cell swelling and eventually causing cell rupture and loss of function, apoptosis is an active process of cell death which also occurs under normal metabolic circumstances.¹⁶ Oxidative stress from the production of excessive amounts of free radicals and nitric oxide and peroxynitrite is involved in noise-related injury.^{8,16,17} Oxidative damage is also associated with medication-induced hearing impairment.¹⁸

In the present study, we investigated the potential preventative effect of NAC in rats exposed to AT. Transmission and scanning electron microscopy evaluations were performed. Our light microscopy and SEM evaluations showed no cochlear damage in group 1 (control group). In group 2 (AT-exposed group), cochlear damage was detected. The inner hair cell stereocilia were missing in some regions, and the OHC stereocilia were severely damaged by noise trauma with a large number prostrate or missing. The damage was more evident in the OHCs in the basal turns of the cochlea. In group 3 (NAC+AT group), although there was cochlear damage, it was less than in group 2 (AT-exposed group without NAC). In group 3, there were fewer missing cells in the inner and OHC group. There was slight damage in the OHC stereocilia.

In light microscopic and SEM evaluations of the control group, the cochlear structure and epithelium were normal. In group 2 (AT group), extensive hair cell loss was observed in the cochlea by light microscopy. In the SEM evaluation, various epithelial damage and loss of stereocilia were also observed. In group 3 (AT+NAC group), light microscopy showed decreased damage with preserved cochlear structures. In the SEM evaluation, although stereocilia loss occurred in this group, nearly normal cell structure and vertical and symmetrical alignment of stereocilia structures were also observed compared to the AT group.

Single agents have demonstrated some preventative benefits against noise-induced damage and medication-related ototoxicity. Vitamin E reduces cochlear damage from these factors.^{19,20} NAC can also protect the inner ear apparatus.^{21,22} Coenzyme Q10 and vitamin C both have shown damage-preventing properties.^{23,24} High-dose NAC has been combined with acetyl-L-carnitine before and after noise exposure to attenuate hearing loss.^{25,26}

The use of NAC has been validated in preventing noise-induced permanent threshold shift (NIPTS).¹⁴ Ewert et al. reported that a combination of antioxidants, 2,4-disulfonyl α -phenyl tertiary butyl nitron (HPN-07) and NAC could enhance temporary threshold shift (TTS) recovery and prevent permanent threshold shift (PTS) by reducing damage to the mechanical and neural components of the auditory system when administered shortly after blast exposure.²⁷

In the present study, NAC reduced the cochlear damage due to AT. Because NAC has antioxidant capacity, it is possible that AT caused increased free radicals and death of OHCs. NAC is an antioxidant agent and prevents cochlear damage due to AT in rats. If it is used before AT, its preventative effects are more obvious. Thus, we recommend the use of NAC in places with high noise exposure.

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